Original Article

Immunotherapy in anaplastic thyroid cancer

Maoguang Ma^{1*}, Bo Lin^{1*}, Mingdian Wang³, Xiaoli Liang¹, Lei Su², Okenwa Okose^{4,5}, Weiming Lv¹, Jie Li^{1,5}

Departments of ¹Breast and Thyroid Surgery, ²Geriatrics, The First Affiliated Hospital of Sun Yat-sen University, Guangzhou, China; ³State Key Laboratory of Oncology in South China and Collaborative Innovation Center for Cancer Medicine, Sun Yat-sen University Cancer Center, Guangzhou, China; ⁴Texas A & M College of Medicine, College Station, TX 77843, USA; ⁵Division of Thyroid and Parathyroid Surgery, Massachusetts Eye and Ear Infirmary, Harvard Medical School, Boston, MA, USA. *Equal contributors.

Received February 12, 2019; Accepted February 13, 2020; Epub March 15, 2020; Published March 30, 2020

Abstract: Anaplastic thyroid cancer (ATC) is one of the worst human malignancies, with an associated median survival of only 5 months. It is resistant to conventional thyroid cancer therapies, including radioiodine and thyroid-stimulating hormone suppression. Cancer immunotherapy has emerged over the past few decades as a transformative approach to treating a wide variety of cancers. However, immunotherapy for ATC is still in the experimental stage. This review will cover several strategies of immunotherapy and discuss the possible application of these strategies in the treatment of ATC (such as targeted therapy for tumor-associated macrophages, cancer vaccines, adoptive immunotherapy, monoclonal antibodies and immune checkpoint blockade) with the hope of improving the prognosis of ATC in the future.

Keywords: Immunotherapy, anaplastic thyroid cancer, immune checkpoint blockade, tumor-associated macrophages, oncolytic virus and neoantigens

Introduction

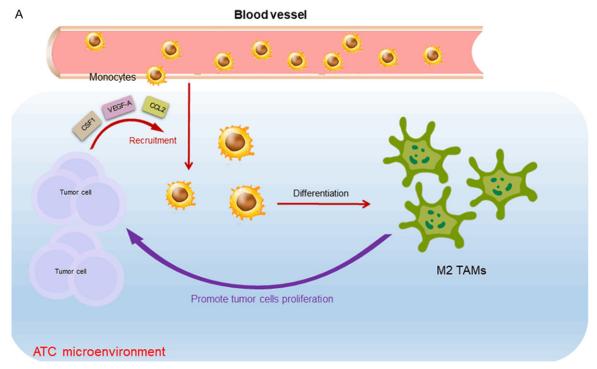
Anaplastic thyroid cancer (ATC) is the most aggressive malignancy among all thyroid cancer subtypes, with a disease-specific mortality approaching 100% [1]. Conventional therapeutics for thyroid cancer include surgery, chemotherapy and radiation treatment, but none of these methods can provide satisfactory efficacy in ATC, so the median survival of patients from diagnosis is still approximately 5 months [2-4].

With the rapid development of and collaborations between oncology, immunology, and molecular biology, great progress has been made in tumor immunotherapy research. James P Allison and Tasuku Honjo were awarded the Nobel Prize in Physiology or Medicine 2018 for their outstanding contribution in promoting immunotherapy in cancer treatment. According to a comprehensive analysis of the clinical immune-oncology landscape, there were more than 2,000 immuno-oncology agents in clinical or preclinical stages until 2017; furthermore, there were 3,042 active clinical trials of these

agents enrolling a total of 577,076 patients [5]. Immunotherapy has become an important means of treatment for many cancers, such as melanoma, non-small-cell lung cancer (NSCLC), and urothelial carcinoma [6-10]. Up to October 2018, the FDA has approved 7 immune checkpoint inhibitors that can be used for a variety of cancer treatments based on the results of related clinical trials [11]. Immunotherapy brings hopes for the treatment of ATC [12, 13]. Here, we reviewed the potential strategies of immunotherapy for ATC, aiming to provide comprehensive information on immunotherapy for this intractable carcinoma.

Targeted therapy for tumor-associated macrophages

Tumor-associated macrophages (TAMs) are derived from circulating monocytes differentiated in the tumor microenvironment [14] (Figure 1A). TAMs have two phenotypes, namely, M1 TAMs, which contribute to immune control over tumors, and M2 TAMs, which promote tumor progression and impair antitumor activity mediated by the immune system [15]. TAMs have been



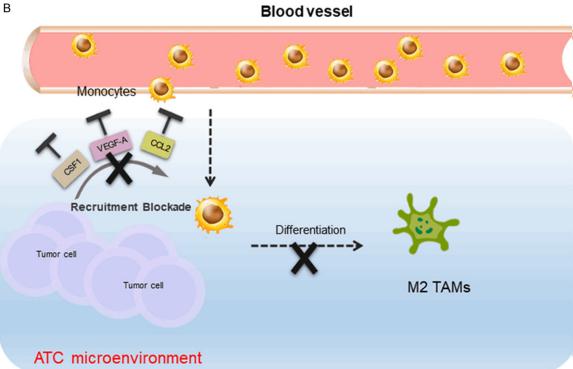


Figure 1. A. Monocytes migrated to tumor microenvironment from blood circulation attracted by CSF1, VEGF-A and CCL2. and differentiated into TAMs (M2 type). M2 TAMs can promote tumor cell proliferation and contribute to disease progress. B. Target therapy for CSF1, VEGF-A and CCL2 to inhibit the recruitment of TAMs from blood circulation, as a result, restricting the pro-tumor function of M2 TAMs and inhibiting tumor cell proliferation.

found to correlate with the immune-suppressive microenvironment in human gliomas and enhance stemness of breast cancer cells [16,

17], which indicates that targeting TAMs is a practical method in gliomas and breast cancer treatment.

In ATC tissues, TAMs represent more than 50% of nucleated cells and are the only type of lymphocyte. All of these TAMs belong to the M2 phenotype [18]. Because a high density of TAMs is closely related to the aggressiveness and invasiveness of ATC, TAMs have been considered a prognostic biomarker for ATC [19, 20]. Considering that TAMs constitute an unusually high proportion of all nucleated cells and form a dense and diffuse network in the tumor microenvironment of ATCs, as identified under electron microscope [18], targeting TAMs may be a feasible way to treat ATCs. Here, we will discuss the strategies for targeting TAMs in ATC.

Inhibiting recruitment of TAMs

During acute inflammatory reactions, monocytes are recruited into tissues from peripheral blood. In this process, chemoattractants and their receptors contribute to the recruitment of these monocytes to inflamed tissues [21, 22]. In the tumor environment, tumor-derived chemoattractants play critical roles in recruiting monocytes into tumors and contribute to the subsequent differentiation of monocytes into M2 TAMs; as such, developing agents to block these chemoattractants has been proposed as a rational way to treat cancer [23] (Figure 1B).

Colony-stimulating factor-1 (CSF1) is a major lineage regulator for most populations of macrophages, playing an important role in recruiting monocytes in many tumors [24-26]. Previous studies have reported that genetic deletion of CSF1 from several models of cancer results in delayed initiation (cervical cancer), progression (breast and pancreatic cancer) and metastasis (breast cancer) associated with the loss of TAMs [27]. CSF1 has been found to be overexpressed in human tissue samples of thyroid tumors, and the expression levels were higher in advanced thyroid cancer than in inactive papillary thyroid cancer (PTC) [28]. In addition, CSF1/CSF1R signaling is required for TAM recruitment and can be pharmacologically targeted to impair PTC initiation [29]. Regarding ATC, the relative expression of CSF1 was higher in ATC cell lines than in PTC cell lines; moreover, both qPCR and microarray data revealed that the expression of CSF1 and its receptor was higher in metastatic ATC cells than in primary ATC cells [30]. These results indicate that the grade of malignancy of thyroid cancer is positively correlated with the expression level of CSF1 and that targeting CSF1/CSF1R signaling may be an effective method in ATC treatment.

Vascular endothelial growth factor A (VEGF-A) can lead to massive infiltration of TAMs into the tumor [31]. Salajegheh A found upregulation of VEGF-A expression in ATC tissues, which indicated that suppression of VEGF-A might also be a potential strategy to inhibit recruitment of TAMs in ATC [32].

Chemokine (C-C motif) ligand 2 (CCL2) is expressed in many cancers, including thyroid cancer [33, 34]. CCL2 can act as a chemotactic factor for monocytes; thus, they can migrate to the tumor region and differentiate into TAMs [35]. P53, a cancer suppressor gene, was reported to play an important role in binding to CCL2 [36]. In addition, p53 mutation was only found in ATC but not in other types of thyroid cancer [37]. It is reasonable to speculate that targeting CCL2 in p53-mutated ATCs may be a promising treatment, but further research is needed to verify this idea.

Due to the critical role of these chemokines in recruiting TAMs, inhibiting chemokine signaling could be considered a therapeutic approach for the treatment of ATC in the future [34].

Repolarization of TAMs from the M2 phenotype to the M1 phenotype

Some scholars argue that tumor tissues have both M1 and M2 TAMs, and the ratio of M1/M2 determines the function of all TAMs. Namely, when the ratio is greater than 1, M1 TAMs accounts for most of the TAMs, and antitumor activity will dominate, in contrast, when the ratio is less than 1, M2 TAMs and thus protumor activity will dominate [38]. Repolarization of M2 TAMs into M1 TAMs has been a widely accepted strategy of immunotherapy for TAMs in ATC [39]. Repolarization is not an independent result of immunotherapy; rather, it is always accompanied by the inhibition of TAM recruitment [15]. Banerjee S demonstrated that heat-killed Mycobacterium indicus pranii (Mw) could induce repolarization of TAMs toward the M1 phenotype in vitro but failed to show this in vivo. Furthermore, Mw combined with a GITR antibody (DTA-1) could repolarize M2 TAMs toward the M1 phenotype in vivo and

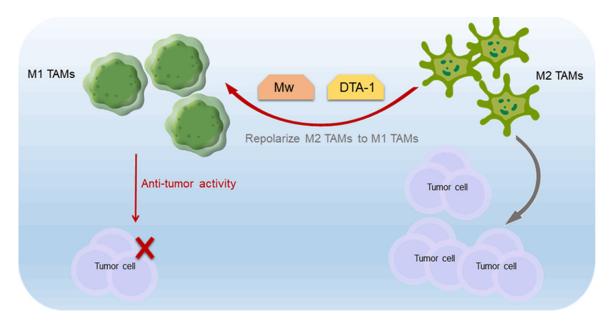


Figure 2. Repolarizing M2 TAMs to M1 TAMs to exert anti-tumor activity and relieving pro-tumor activity, as a result, inhibiting tumor cells progression.

restrict the progression of advanced-stage melanoma [40] (Figure 2). Whether this combination could be used in ATC requires further investigation. Many researchers have found other ways to repolarize M2 TAMs toward the M1 phenotype in ovarian cancer and NSCLC, but none of these means have been applied in an ATC animal model [41, 42]. Since the mechanism of TAM repolarization is still elusive, clarification will require continued efforts.

Cancer vaccines

Cancer vaccines are an important form of immunotherapy. They target defined antigens to induce or expand cancer-specific T cells and rely on DNA, RNA, proteins or peptides [43]. Cancer vaccines can target neoantigens and tumor-associated antigens (TAAs). Neoantigens are uniquely expressed in tumor cells, while TAAs are typically proteins present in normal tissues but overexpressed in cancers [44].

Cancer neoantigens are derived from somatic mutations present in different cancer cells, and these neoantigens are considered significant targets for cancer immunotherapy because of their immunogenicity and lack of expression in normal tissues.

Next-generation sequencing (NGS) can identify the mutational landscape of cancer and is the starting point of cancer neoantigen identifica-

tion [44]. In a recent study, researchers analyzed the genetic alteration patterns of 196 ATC clinical samples and found the two most commonly mutated genes in ATC: TP53 (65%) and TERT (65%). Moreover, after comparing mutation frequencies in ATC and differentiated thyroid cancer (DTC), they identified that their ATC cohort was characterized by an increased frequency of mutations in tumor suppressor genes (TP53, NF2, NF1, and RB1) and PI3K/AKT pathway genes (PIK3CA and PTEN). Furthermore, the authors summarized all the pathways and genes that were altered more frequently in ATC than in differentiated thyroid cancer (DTC) [45]. Their findings provide meaningful genome-level information and can help us to identify neoantigens in ATC. There are many ongoing clinical trials of cancer vaccines for the treatment of different cancers, including breast carcinoma, gastric carcinoma, bladder carcinoma, glioblastoma, kidney cancer, leukemia, lung cancer and melanoma [46].

Peptide/protein platforms are the most common but not the only vaccine platform. Here, we discuss another two types of cancer vaccines that may be used for ATC treatment in the future.

Dendritic cell (DC) vaccines

DCs are potent antigen-presenting cells in the immune system, and mature DCs can elicit

immune stimulation and promote antitumor reactions in the tumor microenvironment [47, 48]. DC vaccines are based on patient-derived DCs armed ex vivo with cognate antigen and costimulatory cues from ligands present on the antigen-presenting cell (APC) surface [49]. Mature DCs loaded with tumor-specific antigen were used in a previous study as a vaccine to treat medullary thyroid carcinoma and proved valid in animal models and some patients [50-53]. Furthermore, a phase I clinical study demonstrated that DC immunotherapy could be administered to thyroid cancer patients without substantial side effects [54]. Landa I reported that ATC had a greater mutation burden than poorly differentiated thyroid cancer and welldifferentiated thyroid cancer [55], suggesting that ATC had a greater chance of harboring tumor neoantigens. Intriguingly, researchers from Argentina found that triiodothyronine could potentiate antitumor responses by bolstering DC-mediated T cell activation during tumor growth in a melanoma mouse model [56], suggesting that DC vaccine-based treatment may be more effective in thyroid cancer than in other tumors due to the high density of triiodothyronine stored in thyroid follicles. The biggest obstacle for applying DC vaccines in ATC is the lack of a tumor-specific antigen at present. With the development of NGS technology and exploration of neoantigens in the future, application of a DC vaccine in the treatment of ATC will be possible.

Oncolytic virus vaccines

Oncolytic virus (OV) therapy is based on the selective replication of viruses in cancer cells and their subsequent spread within a tumor without damage to normal tissue [57]. OVs exert antitumor action through a dual mechanism of selective tumor cell killing and induction of systemic antitumor immunity.

There are two kinds of OVs. The first consists of viruses that naturally replicate preferentially in cancer cells and are nonpathogenic in humans, often due to elevated sensitivity to innate antiviral signaling or dependence on oncogenic signaling pathways. The second consists of viruses that are manipulated genetically for use as vaccine vectors. Until now, many OV therapies have been reported to be effective against ATC cell proliferation, such as dl922-947, Newcastle disease virus, and poxviruses. Interestingly,

dl922-947 has been proven to not only suppress tumor growth but also switch M2 macrophages toward a pro-inflammatory M1 phenotype in an ATC mouse model [58-60].

OVs may break the tolerogenic tumor microenvironment and induce a long-lasting CD8 T cellmediated antitumor response, thereby acting as vaccines [61]. Previous studies have reported that OV infection can counteract cancermediated immune evasion by altering the cytokine milieu and the type of immune cells within the tumor microenvironment; these activities will promote immune-mediated tumor cell recognition and eradication [62, 63]. In addition, the lysis of cancer cells can result in the release of tumor-specific antigens (TSAs) that may have been previously hidden from the immune system because of restricted presentation, consequently triggering powerful antitumor immunity against TSAs [64, 65].

Previous studies have shown that OV vaccines have some efficacy in controlling ATCs both in vivo and in vitro [59, 66]. Based on the results of previous clinical trials, the first oncolytic virus, the HSV-1-based talimogene laherparepvec (T-VEC), was approved for the treatment of nonresectable melanoma in the USA and Europe. Here, we list a collection of clinical trials about the therapeutic effectiveness and safety of OVs in advanced or metastatic solid tumors. ATC patients meet the inclusion criteria of all these clinical trials (Table 1).

Adoptive immunotherapy

Adoptive cell transfer (ACT) is an immunotherapy that relies on the active in vivo recruitment of sufficient numbers of antitumor T cells with the functions necessary to mediate cancer regression [67]. There are two kinds of ACT methods: the first type uses natural host cells that are expanded ex vivo and infused into patients with malignant diseases, and the second method uses infused autologous T cells that have been genetically engineered with chimeric antigen receptors (CARs), which enable these T cells to specifically identify and kill tumor cells to treat different malignant diseases [68, 69].

CAR-T cells can recognize TAAs and induce strong antitumor activity, which has been demonstrated in many cohorts suffering from he-

Immunotherapy for ATC

Table 1. Clinical trials about OVs in the treatment of advanced/metastasis solid tumor

Trial	Condition/Disease	Intervention	Phase	Estimated/Actual Enrollment	State of Trial
NCT00794131	Advanced solid tumor	GL-ONC1	1	43	Completed
NCT02428036	Solid tumors with superficial lesions	TBI-1401	1	6	Completed
NCT03866525	Advanced solid tumor	OH2	1	150	Not yet recruiting
NCT01598129	Advanced solid tumor	ONCOS-102+cyclophosphamide	1	12	Completed
NCT03889275	Advanced solid tumor	MEDI5395+Durvalumab	1	164	Not yet recruiting
NCT02045602	Advanced solid tumor	VCN-01 with or without Abraxane®/Gemcitabine	1	36	recruiting
NCT00625456	Advanced solid tumor	Recombinant Vaccinia GM-CSF; RAC VAC GM-CSF (JX-594)	I	23	Completed

matological tumors, such as CD-19-expressing B cell acute lymphocytic leukemia (B-ALL), chronic lymphocytic leukemia (CLL) and B cell non-Hodgkin lymphoma (NHL) [70, 71]. Many studies have shown that CAR-T cells targeting intercellular adhesion molecule-1 (ICAM-1) can exhibit impressive therapeutic efficacy and survival benefits in a mouse model of ATC [31, 72, 73]. This result indicates that ICAM-1 may be a promising target for CAR-T cell treatment for ATC. Identifying neoantigens is the primary task for the development and application of ACT in ATC treatment.

Monoclonal antibody

Monoclonal antibodies (mAbs) are antibodies produced by identical immune cells that are all clones of a unique parent cell, and mAbs can target tumor cells specifically by engaging surface antigens expressed in cancers; the interaction of mAbs with antigens induces cellular events, such as apoptosis [74]. mAbs can be used alone or conjugated with other agents, and the antitumor effect will be enhanced by this conjugation [75, 76]. This method was deemed an ideal cancer treatment model because it directly targets malignant cells and exerts little cytotoxic effects on noncancerous cells.

Aberrant lipid metabolism in ATCs has drawn the attention of many researchers in recent years. Copland JA revealed that stearoyl-CoA desaturase 1 (SCD1), an important component in de novo lipid biosynthesis, is overexpressed in ATCs and is critical for ATC cell survival and proliferation, which suggests SCD1 as a novel therapeutic target for ATCs [77]. Moreover, gene array analysis of ATC tissue compared to normal thyroid tissue demonstrated increased expression of the machinery that facilitates de novo fatty acid biosynthesis, including acetyl-CoA carboxylase (ACC), fatty acid synthase (FASN) and a variety of proteins whose roles include fatty acid uptake, transport and metabolism [78].

mAbs have been used for many cancer treatments, such as the rituximab target CD20 in non-Hodgkin B cell lymphoma, the cetuximab target EGFR in NSCLC and the trastuzumab target HER2 in breast cancer [79, 80]. The FDA has approved many mAbs for cancer treatment [75]. The obstacle to applying mAbs in ATC is

the current lack of recognized tumor-specific antigens. Advances in NGS technologies have made it possible to compare tumor and normal sequences rapidly and cost-effectively, which provides tremendous help for researchers to explore neoantigens in ATC.

Immune checkpoint blockade

The induction of an immune checkpoint receptor on cancer cells could evade immune attack, resulting in tumor proliferation, invasion, and metastasis [81, 82]. PD-1 and CTLA-4 are the most well-studied immune checkpoint receptors at present (Figure 3).

Programmed cell death protein 1 (PD-1) blockade

PD-1 inhibitors have been used to treat many cancers, such as melanoma, NSCLC, and renal carcinoma [83-85]. Owing to their efficacy in suppressing tumor growth and inducing tumor cell apoptosis, the Food and Drug Administration has approved many PD-1 inhibitors for the treatment of a variety of cancers, such as pembrolizumab for melanoma and nivolumab for NSCLC [86, 87]. No such drug has been approved for ATC due to a lack of evidence from clinical trials.

PD-L1, the ligand of PD-1, is expressed in 28.6% of ATC patients, a percentage that is higher than that observed in well-differentiated thyroid cancer [88]. The latest statistics from Cantara indicate that the PD-L1 positive rate in ATCs reaches up to 70-90%, and PD-L1 mAb treatment can reduce tumor volume in an ATC mouse model [89]. In a recent study, Goodman AM reported that PD-L1 copy number alterations were found in a small subgroup of diverse solid tumors and may correlate with responses to checkpoint blockade. His data showed that thyroid anaplastic carcinoma has a relatively high frequency of PD-L1 amplification among most solid tumors, which means that ATCs may be more sensitive to PD-L1 inhibitors than solid tumors with a lower frequency of PD-L1 amplification [90].

Brauner E revealed that the expression of PD-L1 in ATCs was correlated with the BRAFV600E mutation; in addition, anti-PD-L1 treatment potentiated the effect of a BRAF inhibitor. Furthermore, the tumor volume was

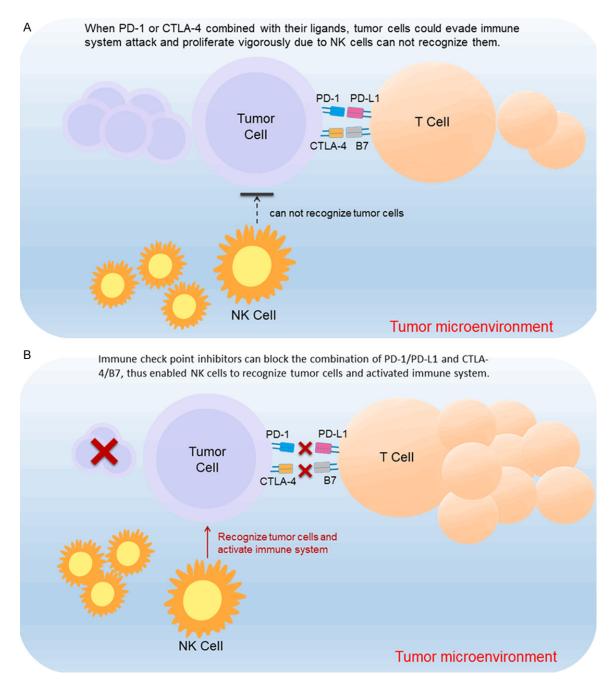


Figure 3. A. Several immune checkpoints are overexpressed on T cell surface in anaplastic thyroid cancer microenvironment and their corresponding ligands on tumor cells. Interaction of receptors and ligands lead to inhibition of T cell proliferation and progression of tumor cells. B. Immune checkpoints inhibitor prevent the combination of immune checkpoints and their ligands, destroying the immune escape mechanism of tumor cell and enhancing anti-tumor activity of NK cells.

decreased more strikingly by the combination of these two therapies than by either of these two agents used alone in an ATC mouse model (the tumor volume of the combination group was reduced by as much as 81% compared to that of the placebo group, while it was reduced by 8% and 44% for the PD-L1 mAb group and

BRAF inhibitor group, respectively) [91]. Similar therapeutic effectiveness was observed in some clinical cases [92, 93]. Another study has shown that the ratio of CD56^{hi}CD16^{hi/lo} NK cells to all NK cells is higher in the peripheral blood of ATC patients than in patients with other types of thyroid cancer, while the CD56^{hi}CD16^{hi/lo} NK

Immunotherapy for ATC

Table 2. Ongoing clinical trials about PD-1/PD-L1 inhibitor used for ATC patients

Trial	Intervention	Phase	Estimated enrollment	Estimated completion date (month/year)
	durvalumab			
NCT03122497	tremelimumab	1	12	5/2020
	SBRT			
NCT02688608	Pembrolizumab	II	20	10/2020
	Pembrolizumab			
	Chemotherapy			
NCT03211117	IMRT	II	3	12/2019
	With or without surgery			
NCT02936102	PDR001	1	155 (Comprise a cohort of ATC patients)	2/2020
	FAZ053			
NCT03181100	Atezolizumab	II	50	7/2023
	Chemotherapy			
NCT03246958	Nivolumab	II	54	3/2025
	Ipilimumab			
NCT02404441	PDR001	II	319 (Comprise a cohort of ATC patients)	5/2020

SBRT: Stereo-tactic Body Radiation Therapy; IMRT: Intensity-Modulated Radiation Therapy.

cells presented higher PD-1 than other kinds of NK cells [94]. Taken together, this information suggests that a PD-L1 inhibitor is a promising treatment for ATC. A list of ongoing clinical trials pertaining to the potential efficacy of this agent for ATC is shown here (**Table 2**).

Cytotoxic T lymphocyte antigen 4 (CTLA-4) blockade

CTLA-4 is another immune checkpoint, also known as CD152. Previous studies found that CTLA-4 is primarily involved in the regulation of T cell activation in lymph nodes and in Tregmediated suppression of DC activity. A recent study showed that CTLA-4 binding to its ligands could shield cancer cells from cytotoxic T lymphocyte-mediated attack [95]. Anti-CTLA-4 antibodies have shown promising results in cancer treatment, and three antibodies, ipilimumab, tremelimumab and MK1308, have been largely involved in clinical trials [96, 97]. However, one recent study showed that CD80, a ligand of CTLA-4, was downregulated in 9 out of 11 ATC patients [98]. Whether CTLA-4 exerts an immunosuppressive function in ATCs by binding to its ligands remains elusive. Two trials (NCT03122497 and NCT03246958) contained a cohort of ATCs treated with CTLA-4 inhibitors.

In addition, ATC had strikingly more genetic alterations per tumor than any other thyroid cancer subtype [46, 55]. Furthermore, mutation burden is strongly correlated with favorable clinical benefit of checkpoint blockade

therapy [99, 100]. From these data provided by other researchers, we surmise that ATC patients are very likely to benefit from immune checkpoint inhibitors and look forward to the outcome of ongoing clinical trials concerning this therapeutic method.

Future perspective

ATC is still a challenge for clinical experts due to its dedifferentiated phenotype and aggressive features. Although immunotherapy has been shown as a promising strategy for this intractable cancer, there are many concerns regarding this treatment that need to be resolved.

Response rates to one single method of immunotherapy are not satisfactory in many cancers, and crosstalk between different immunotherapies has been observed in previous studies; for example, in addition to directly assaulting cancer cells, OVs can also cause repolarization of M2 macrophages to M1 macrophages, decreasing TAM density [65]. In addition, TAMs can express cytokines and enzymes that can suppress T cell recruitment and activation, thereby promoting resistance to immune checkpoint inhibition [101]. Researchers have demonstrated that the combination of a CSF1R inhibitor with a CXCR2 inhibitor can significantly reduce tumor growth; moreover, when a PD-1 antibody was added to this combination, it resulted in blockade of tumor growth [102]. We speculate that different immunotherapies may interact reciprocally rather than independently of one another, illuminating the combination of a variety of different strategies of immunotherapy as a new direction in the future that may enhance antitumor efficacy and contribute to a better prognosis of ATC.

Many immunotherapy methods, such as tumor vaccines, ACT, and mAbs, rely on targeting tumor-associated antigens (TAAs) or neoantigens presented on cancer cells. To date, no single TAA or neoantigen for ATC has been proven valid, but there are a couple of candidates that might become the first, such as ICAM-1, CD47, CD70, autotoxin and CD1d [103-105]. Recent advances in NGS and epitope prediction have made the rapid identification of tumor neoantigens possible. In the future, detecting TAAs and neoantigens will be critical for the development of immunotherapy in ATC.

OVs are also very impressive for their ability to inhibit ATC proliferation and progression, although the full mechanism of antitumor action remains elusive. Moreover, the reason why these viruses target cancer cells and do not attack normal tissues is still unknown. Is there a connection between TAAs and the OV? Further investigations are needed to provide an explanation of this phenomenon. Nevertheless, OVs are a highly promising approach for cancer treatment, and illuminating the antitumor mechanism of OVs may guide us to exploit new immunotherapy methods for cancer treatment.

Despite the tremendous success of immune checkpoint inhibitors, there are many patients who still do not respond to immunotherapy or develop therapeutic resistance [106-108]. Insufficient immune activation is considered one of the main reasons for low response rates, and a combination of checkpoint blockers has been proposed to increase the response rates [109]. Furthermore, Ishizuka JJ reported that the ADAR1 gene was critical for resistance to immunotherapy and proved in a melanoma mouse model that loss of ADAR1 could overcome resistance to immunotherapy. In addition, their results showed that loss of function of ADAR1 restored sensitivity to immunotherapy in tumors with a B2m deletion, which means that targeting the ADAR1 gene is an effective immunotherapy strategy even in the absence of a tumor-specific endogenous CD8+ T cell response [110]. There are no immunocytes in the microenvironment of ATCs except M2 macrophages. This seems to be a disadvantage for immunotherapy, but research provides another avenue for immunotherapy, and more importantly, immunotherapy is likely suitable for ATC given the tumor microenvironment of this unique disease (featuring a lack of a tumorspecific endogenous CD8+ T cell response).

Conclusion

Immunotherapy has demonstrated excellent efficacy for some malignancies, such as melanoma, NSCLC, and leukemia. As effective treatments for ATC are still limited, it is urgent to explore new therapies for this untreatable disease. Generally, immunotherapy has not been approved for ATC, but it has shown palpable efficacy in some animal models of ATC. As a promising means for controlling ATC, more attention should be given to the prompt development of future immunotherapies.

Acknowledgements

This work was supported by the Guangdong Science and Technology Department Fund: 2016A040403049; 2017A010105029.

Disclosure of conflict of interest

None.

Address correspondence to: Weiming Lv and Jie Li, Department of Thyroid and Breast Surgery, The First Affiliated Hospital, Sun Yat-sen University, Guangzhou 510000, China. Tel: +86-13925177549; Fax: +86-20-87755766-8198; E-mail: lvwm@mail.sysu.edu.cn (WML); Tel: +86-13602790063; Fax: +86-20-87755766-8198; E-mail: lijie78@mail.sysu.edu.cn (JL)

References

- [1] Are C and Shaha AR. Anaplastic thyroid carcinoma: biology, pathogenesis, prognostic factors, and treatment approaches. Ann Surg Oncol 2006; 13: 453-464.
- [2] Hsu KT, Yu XM, Audhya AW, Jaume JC, Lloyd RV, Miyamoto S, Prolla TA and Chen H. Novel approaches in anaplastic thyroid cancer therapy. Oncologist 2014; 19: 1148-1155.
- [3] Smallridge RC, Ain KB, Asa SL, Bible KC, Brierley JD, Burman KD, Kebebew E, Lee NY, Nikiforov YE, Rosenthal MS, Shah MH, Shaha AR and Tuttle RM. American thyroid association guidelines for management of patients with anaplastic thyroid cancer. Thyroid 2012; 22: 1104-1139.
- [4] Untch BR and Olson JA Jr. Anaplastic thyroid carcinoma, thyroid lymphoma, and metastasis

- to thyroid. Surg Oncol Clin N Am 2006; 15: 661-679.
- [5] Tang J, Shalabi A and Hubbard-Lucey VM. Comprehensive analysis of the clinical immuno-oncology landscape. Ann Oncol 2018; 29: 84-91.
- [6] Rusch T, Bayry J, Werner J, Shevchenko I and Bazhin AV. Immunotherapy as an option for cancer treatment. Arch Immunol Ther Exp (Warsz) 2018; 66: 89-96.
- [7] Rodriguez-Cerdeira C, Carnero Gregorio M, Lopez-Barcenas A, Sanchez-Blanco E, Sanchez-Blanco B, Fabbrocini G, Bardhi B, Sinani A and Guzman RA. Advances in immunotherapy for melanoma: a comprehensive review. Mediators Inflamm 2017; 2017: 3264217.
- [8] Mayor M, Yang N, Sterman D, Jones DR and Adusumilli PS. Immunotherapy for non-small cell lung cancer: current concepts and clinical trials. Eur J Cardiothorac Surg 2016; 49: 1324-1333.
- [9] Carlo MI, Voss MH and Motzer RJ. Checkpoint inhibitors and other novel immunotherapies for advanced renal cell carcinoma. Nat Rev Urol 2016; 13: 420-431.
- [10] Suzman DL, Agrawal S, Ning YM, Maher VE, Fernandes LL, Karuri S, Tang S, Sridhara R, Schroeder J, Goldberg KB, Ibrahim A, McKee AE, Pazdur R and Beaver JA. FDA approval summary: atezolizumab or pembrolizumab for the treatment of patients with advanced urothelial carcinoma ineligible for cisplatin-containing chemotherapy. Oncologist 2019; 24: 563-569.
- [11] Li Z, Song W, Rubinstein M and Liu D. Recent updates in cancer immunotherapy: a comprehensive review and perspective of the 2018 China Cancer Immunotherapy Workshop in Beijing. J Hematol Oncol 2018; 11: 142.
- [12] Cabanillas ME, Ryder M and Jimenez C. Targeted therapy for advanced thyroid cancer: kinase inhibitors and beyond. Endocr Rev 2019; 40: 1573-1604.
- [13] Saini S, Tulla K, Maker AV, Burman KD and Prabhakar BS. Therapeutic advances in anaplastic thyroid cancer: a current perspective. Mol Cancer 2018; 17: 154.
- [14] Tang X, Mo C, Wang Y, Wei D and Xiao H. Antitumour strategies aiming to target tumour-associated macrophages. Immunology 2013; 138: 93-104.
- [15] Naoum GE, Morkos M, Kim B and Arafat W. Novel targeted therapies and immunotherapy for advanced thyroid cancers. Mol Cancer 2018; 17: 51.
- [16] Pinton L, Masetto E, Vettore M, Solito S, Magri S, D'Andolfi M, Del Bianco P, Lollo G, Benoit JP, Okada H, Diaz A, Della Puppa A and Mandruzzato S. The immune suppressive microenvironment of human gliomas depends on the accumulation of bone marrow-derived macrophages

- in the center of the lesion. J Immunother Cancer 2019; 7: 58.
- [17] Liu D, Lu Q, Wang X, Wang J, Lu N, Jiang Z, Hao X, Li J, Liu J, Cao P, Peng G, Tao Y, Zhao D, He F and Tang L. LSECtin on tumor-associated macrophages enhances breast cancer stemness via interaction with its receptor BTN3A3. Cell Res 2019; 29: 365-378.
- [18] Caillou B, Talbot M, Weyemi U, Pioche-Durieu C, Al Ghuzlan A, Bidart JM, Chouaib S, Schlumberger M and Dupuy C. Tumor-associated macrophages (TAMs) form an interconnected cellular supportive network in anaplastic thyroid carcinoma. PLoS One 2011; 6: e22567.
- [19] Kim DI, Kim E, Kim YA, Cho SW, Lim JA and Park YJ. Macrophage densities correlated with CXC chemokine receptor 4 expression and related with poor survival in anaplastic thyroid cancer. Endocrinol Metab (Seoul) 2016; 31: 469-475.
- [20] Ryder M, Ghossein RA, Ricarte-Filho JC, Knauf JA and Fagin JA. Increased density of tumorassociated macrophages is associated with decreased survival in advanced thyroid cancer. Endocr Relat Cancer 2008; 15: 1069-1074.
- [21] Schmid MC, Franco I, Kang SW, Hirsch E, Quilliam LA and Varner JA. PI3-kinase gamma promotes Rap1a-mediated activation of myeloid cell integrin alpha4beta1, leading to tumor inflammation and growth. PLoS One 2013; 8: e60226.
- [22] Cai Z, Chen Q, Chen J, Lu Y, Xiao G, Wu Z, Zhou Q and Zhang J. Monocyte chemotactic protein 1 promotes lung cancer-induced bone resorptive lesions in vivo. Neoplasia 2009; 11: 228-236.
- [23] Kaneda MM, Cappello P, Nguyen AV, Ralainirina N, Hardamon CR, Foubert P, Schmid MC, Sun P, Mose E, Bouvet M, Lowy AM, Valasek MA, Sasik R, Novelli F, Hirsch E and Varner JA. Macrophage Pl3Kgamma drives pancreatic ductal adenocarcinoma progression. Cancer Discov 2016; 6: 870-885.
- [24] Cannarile MA, Weisser M, Jacob W, Jegg AM, Ries CH and Rüttinger D. Colony-stimulating factor 1 receptor (CSF1R) inhibitors in cancer therapy. J Immunother Cancer 2017; 5: 53.
- [25] Neubert NJ, Schmittnaegel M, Bordry N, Nassiri S, Wald N, Martignier C, Tille L, Homicsko K, Damsky W, Maby-El Hajjami H, Klaman I, Danenberg E, Ioannidou K, Kandalaft L, Coukos G, Hoves S, Ries CH, Fuertes Marraco SA, Foukas PG, De Palma M and Speiser DE. T cell-induced CSF1 promotes melanoma resistance to PD1 blockade. Sci Transl Med 2018; 10.
- [26] Edwards DKt, Watanabe-Smith K, Rofelty A, Damnernsawad A, Laderas T, Lamble A, Lind EF, Kaempf A, Mori M, Rosenberg M, d'Almeida A, Long N, Agarwal A, Sweeney DT, Loriaux M, McWeeney SK and Tyner JW. CSF1R inhibitors

- exhibit antitumor activity in acute myeloid leukemia by blocking paracrine signals from support cells. Blood 2019; 133: 588-599.
- [27] Noy R and Pollard JW. Tumor-associated macrophages: from mechanisms to therapy. Immunity 2014; 41: 49-61.
- [28] Kim S, Cho SW, Min HS, Kim KM, Yeom GJ, Kim EY, Lee KE, Yun YG, Park DJ and Park YJ. The expression of tumor-associated macrophages in papillary thyroid carcinoma. Endocrinol Metab (Seoul) 2013; 28: 192-198.
- [29] Ryder M, Gild M, Hohl TM, Pamer E, Knauf J, Ghossein R, Joyce JA and Fagin JA. Genetic and pharmacological targeting of CSF-1/CSF-1R inhibits tumor-associated macrophages and impairs BRAF-induced thyroid cancer progression. PLoS One 2013; 8: e54302.
- [30] Garg M, Okamoto R, Nagata Y, Kanojia D, Venkatesan S, M TA, Braunstein GD, Said JW, Doan NB, Ho Q, Akagi T, Gery S, Liu LZ, Tan KT, Chng WJ, Yang H, Ogawa S and Koeffler HP. Establishment and characterization of novel human primary and metastatic anaplastic thyroid cancer cell lines and their genomic evolution over a year as a primagraft. J Clin Endocrinol Metab 2015; 100: 725-735.
- [31] Min IM, Shevlin E, Vedvyas Y, Zaman M, Wyrwas B, Scognamiglio T, Moore MD, Wang W, Park S, Park S, Panjwani S, Gray KD, Tassler AB, Zarnegar R, Fahey TJ 3rd and Jin MM. CAR T therapy targeting ICAM-1 eliminates advanced human thyroid tumors. Clin Cancer Res 2017; 23: 7569-7583.
- [32] Salajegheh A, Pakneshan S, Rahman A, Dolan-Evans E, Zhang S, Kwong E, Gopalan V, Lo CY, Smith RA and Lam AK. Co-regulatory potential of vascular endothelial growth factor-A and vascular endothelial growth factor-C in thyroid carcinoma. Hum Pathol 2013; 44: 2204-2212.
- [33] Ferrari SM, Elia G, Piaggi S, Baldini E, Ulisse S, Miccoli M, Materazzi G, Antonelli A and Fallahi P. CCL2 is modulated by cytokines and PPARgamma in anaplastic thyroid cancer. Anticancer Agents Med Chem 2018; 18: 458-466.
- [34] Antonelli A, Ferrari SM and Fallahi P. Current and future immunotherapies for thyroid cancer. Expert Rev Anticancer Ther 2018; 18: 149-159.
- [35] Richards DM, Hettinger J and Feuerer M. Monocytes and macrophages in cancer: development and functions. Cancer Microenviron 2013; 6: 179-191.
- [36] Tang X and Amar S. p53 suppresses CCL2-induced subcutaneous tumor xenograft. Tumour Biol 2015; 36: 2801-2808.
- [37] Zhu X, Zhao L, Park JW, Willingham MC and Cheng SY. Synergistic signaling of KRAS and thyroid hormone receptor beta mutants promotes undifferentiated thyroid cancer through

- MYC up-regulation. Neoplasia 2014; 16: 757-769.
- [38] Mantovani A, Marchesi F, Malesci A, Laghi L and Allavena P. Tumour-associated macrophages as treatment targets in oncology. Nat Rev Clin Oncol 2017; 14: 399-416.
- [39] Parayath NN, Parikh A and Amiji MM. Repolarization of tumor-associated macrophages in a genetically engineered nonsmall cell lung cancer model by intraperitoneal administration of hyaluronic acid-based nanoparticles encapsulating microRNA-125b. Nano Lett 2018; 18: 3571-3579.
- [40] Banerjee S, Halder K, Ghosh S, Bose A and Majumdar S. The combination of a novel immunomodulator with a regulatory T cell suppressing antibody (DTA-1) regress advanced stage B16F10 solid tumor by repolarizing tumor associated macrophages in situ. Oncoimmunology 2015; 4: e995559.
- [41] Lizotte PH, Baird JR, Stevens CA, Lauer P, Green WR, Brockstedt DG and Fiering SN. Attenuated Listeria monocytogenes reprograms M2-polarized tumor-associated macrophages in ovarian cancer leading to iNOS-mediated tumor cell lysis. Oncoimmunology 2014; 3: e28926.
- [42] Downey CM, Aghaei M, Schwendener RA and Jirik FR. DMXAA causes tumor site-specific vascular disruption in murine non-small cell lung cancer, and like the endogenous non-canonical cyclic dinucleotide STING agonist, 2'3'-cGAMP, induces M2 macrophage repolarization. PLoS One 2014; 9: e99988.
- [43] Calvo Tardon M, Allard M, Dutoit V, Dietrich PY and Walker PR. Peptides as cancer vaccines. Curr Opin Pharmacol 2019; 47: 20-26.
- [44] Li L, Goedegebuure SP and Gillanders WE. Preclinical and clinical development of neoantigen vaccines. Ann Oncol 2017; 28: xii11-xii17.
- [45] Pozdeyev N, Gay LM, Sokol ES, Hartmaier R, Deaver KE, Davis S, French JD, Borre PV, La-Barbera DV, Tan AC, Schweppe RE, Fishbein L, Ross JS, Haugen BR and Bowles DW. Genetic analysis of 779 advanced differentiated and anaplastic thyroid cancers. Clin Cancer Res 2018; 24: 3059-3068.
- [46] Bezu L, Kepp O, Cerrato G, Pol J, Fucikova J, Spisek R, Zitvogel L, Kroemer G and Galluzzi L. Trial watch: peptide-based vaccines in anticancer therapy. Oncoimmunology 2018; 7: e1511506.
- [47] Ugolini C, Basolo F, Proietti A, Vitti P, Elisei R, Miccoli P and Toniolo A. Lymphocyte and immature dendritic cell infiltrates in differentiated, poorly differentiated, and undifferentiated thyroid carcinoma. Thyroid 2007; 17: 389-393.
- [48] Elster JD, Krishnadas DK and Lucas KG. Dendritic cell vaccines: a review of recent developments and their potential pediatric applica-

- tion. Hum Vaccin Immunother 2016; 12: 2232-2239.
- [49] Bol KF, Schreibelt G, Gerritsen WR, de Vries IJ and Figdor CG. Dendritic cell-based immunotherapy: state of the art and beyond. Clin Cancer Res 2016; 22: 1897-1906.
- [50] Schott M, Seissler J, Lettmann M, Fouxon V, Scherbaum WA and Feldkamp J. Immunotherapy for medullary thyroid carcinoma by dendritic cell vaccination. J Clin Endocrinol Metab 2001; 86: 4965-4969.
- [51] Bachleitner-Hofmann T, Friedl J, Hassler M, Hayden H, Dubsky P, Sachet M, Rieder E, Pfragner R, Brostjan C, Riss S, Niederle B, Gnant M and Stift A. Pilot trial of autologous dendritic cells loaded with tumor lysate(s) from allogeneic tumor cell lines in patients with metastatic medullary thyroid carcinoma. Oncol Rep 2009; 21: 1585-1592.
- [52] Papewalis C, Wuttke M, Seissler J, Meyer Y, Kessler C, Jacobs B, Ullrich E, Willenberg HS, Schinner S, Baehring T, Scherbaum WA and Schott M. Dendritic cell vaccination with xenogenic polypeptide hormone induces tumor rejection in neuroendocrine cancer. Clin Cancer Res 2008; 14: 4298-4305.
- [53] Stift A, Sachet M, Yagubian R, Bittermann C, Dubsky P, Brostjan C, Pfragner R, Niederle B, Jakesz R, Gnant M and Friedl J. Dendritic cell vaccination in medullary thyroid carcinoma. Clin Cancer Res 2004; 10: 2944-2953.
- [54] Kuwabara K, Nishishita T, Morishita M, Oyaizu N, Yamashita S, Kanematsu T, Obara T, Mimura Y, Inoue Y, Kaminishi M, Kaga K, Amino N, Kitaoka M, Ito K, Miyauchi A, Noguchi S, Uchimaru K, Akagawa E, Watanabe N, Takahashi TA, Sato K, Inazawa T, Nakaoka T and Yamashita N. Results of a phase I clinical study using dendritic cell vaccinations for thyroid cancer. Thyroid 2007; 17: 53-58.
- [55] Landa I, Ibrahimpasic T, Boucai L, Sinha R, Knauf JA, Shah RH, Dogan S, Ricarte-Filho JC, Krishnamoorthy GP, Xu B, Schultz N, Berger MF, Sander C, Taylor BS, Ghossein R, Ganly I and Fagin JA. Genomic and transcriptomic hallmarks of poorly differentiated and anaplastic thyroid cancers. J Clin Invest 2016; 126: 1052-1066.
- [56] Alamino VA, Montesinos MM, Rabinovich GA and Pellizas CG. The thyroid hormone triiodothyronine reinvigorates dendritic cells and potentiates anti-tumor immunity. Oncoimmunology 2016; 5: e1064579.
- [57] Russell SJ, Peng KW and Bell JC. Oncolytic virotherapy. Nat Biotechnol 2012; 30: 658-670.
- [58] Mundi N, Um S, Yoo J, Rizzo G, Black M, Pinto N, Palma DA, Fung K, MacNeil D, Mymryk JS, Barrett JW and Nichols AC. The control of anaplastic thyroid carcinoma cell lines by oncolytic poxviruses. Virus Res 2014; 190: 53-59.

- [59] Passaro C, Borriello F, Vastolo V, Di Somma S, Scamardella E, Gigantino V, Franco R, Marone G and Portella G. The oncolytic virus dl922-947 reduces IL-8/CXCL8 and MCP-1/CCL2 expression and impairs angiogenesis and macrophage infiltration in anaplastic thyroid carcinoma. Oncotarget 2016; 7: 1500-1515.
- [60] Jiang K, Song C, Kong L, Hu L, Lin G, Ye T, Yao G, Wang Y, Chen H, Cheng W, Barr MP, Liu Q, Zhang G, Ding C and Meng S. Recombinant oncolytic Newcastle disease virus displays antitumor activities in anaplastic thyroid cancer cells. BMC Cancer 2018; 18: 746.
- [61] Bartlett DL, Liu Z, Sathaiah M, Ravindranathan R, Guo Z, He Y and Guo ZS. Oncolytic viruses as therapeutic cancer vaccines. Mol Cancer 2013: 12: 103.
- [62] Prestwich RJ, Errington F, Diaz RM, Pandha HS, Harrington KJ, Melcher AA and Vile RG. The case of oncolytic viruses versus the immune system: waiting on the judgment of Solomon. Hum Gene Ther 2009; 20: 1119-1132.
- [63] Di Paolo NC, Miao EA, Iwakura Y, Murali-Krishna K, Aderem A, Flavell RA, Papayannopoulou T and Shayakhmetov DM. Virus binding to a plasma membrane receptor triggers interleukin-1 alpha-mediated proinflammatory macrophage response in vivo. Immunity 2009; 31: 110-121.
- [64] Kaufman HL, Kohlhapp FJ and Zloza A. Oncolytic viruses: a new class of immunotherapy drugs. Nat Rev Drug Discov 2015; 14: 642-662.
- [65] Chiocca EA and Rabkin SD. Oncolytic viruses and their application to cancer immunotherapy. Cancer Immunol Res 2014; 2: 295-300.
- [66] Lin SF, Price DL, Chen CH, Brader P, Li S, Gonzalez L, Zhang Q, Yu YA, Chen N, Szalay AA, Fong Y and Wong RJ. Oncolytic vaccinia virotherapy of anaplastic thyroid cancer in vivo. J Clin Endocrinol Metab 2008; 93: 4403-4407.
- [67] Ruella M and Kalos M. Adoptive immunotherapy for cancer. Immunol Rev 2014; 257: 14-38.
- [68] Rosenberg SA and Restifo NP. Adoptive cell transfer as personalized immunotherapy for human cancer. Science 2015; 348: 62-68.
- [69] Wang L, Yao R, Zhang L, Fan C, Ma L and Liu J. Chimeric antigen receptor T cell therapy and other therapeutics for malignancies: combination and opportunity. Int Immunopharmacol 2019; 70: 498-503.
- [70] Wang Z, Wu Z, Liu Y and Han W. New development in CAR-T cell therapy. J Hematol Oncol 2017; 10: 53.
- [71] Maude SL, Frey N, Shaw PA, Aplenc R, Barrett DM, Bunin NJ, Chew A, Gonzalez VE, Zheng Z, Lacey SF, Mahnke YD, Melenhorst JJ, Rheingold SR, Shen A, Teachey DT, Levine BL, June CH, Porter DL and Grupp SA. Chimeric antigen receptor T cells for sustained remissions in

- leukemia. N Engl J Med 2014; 371: 1507-1517.
- [72] Vedvyas Y, Shevlin E, Zaman M, Min IM, Amor-Coarasa A, Park S, Park S, Kwon KW, Smith T, Luo Y, Kim D, Kim Y, Law B, Ting R, Babich J and Jin MM. Longitudinal PET imaging demonstrates biphasic CAR T cell responses in survivors. JCl Insight 2016; 1: e90064.
- [73] Vedvyas Y, McCloskey JE, Yang Y, Min IM, Fahey TJ, Zarnegar R, Hsu YS, Hsu JM, Besien KV, Gaudet I, Law P, Kim NJ, Hofe EV and Jin MM. Manufacturing and preclinical validation of CAR T cells targeting ICAM-1 for advanced thyroid cancer therapy. Sci Rep 2019; 9: 10634.
- [74] Weiner GJ. Building better monoclonal antibody-based therapeutics. Nat Rev Cancer 2015: 15: 361-370.
- [75] Gharwan H and Groninger H. Kinase inhibitors and monoclonal antibodies in oncology: clinical implications. Nat Rev Clin Oncol 2016; 13: 209-227.
- [76] Birrer MJ, Moore KN, Betella I and Bates RC. Antibody-drug conjugate-based therapeutics: state of the science. J Natl Cancer Inst 2019; 111: 538-549.
- [77] von Roemeling CA, Marlow LA, Pinkerton AB, Crist A, Miller J, Tun HW, Smallridge RC and Copland JA. Aberrant lipid metabolism in anaplastic thyroid carcinoma reveals stearoyl CoA desaturase 1 as a novel therapeutic target. J Clin Endocrinol Metab 2015; 100: E697-709.
- [78] von Roemeling CA and Copland JA. Targeting lipid metabolism for the treatment of anaplastic thyroid carcinoma. Expert Opin Ther Targets 2016; 20: 159-166.
- [79] Le DT, Pardoll DM and Jaffee EM. Cellular vaccine approaches. Cancer J 2010; 16: 304-310.
- [80] Silva AP, Coelho PV, Anazetti M and Simioni PU. Targeted therapies for the treatment of nonsmall-cell lung cancer: monoclonal antibodies and biological inhibitors. Hum Vaccin Immunother 2017; 13: 843-853.
- [81] McDermott J and Jimeno A. Pembrolizumab: PD-1 inhibition as a therapeutic strategy in cancer. Drugs Today (Barc) 2015; 51: 7-20.
- [82] Topalian SL, Taube JM, Anders RA and Pardoll DM. Mechanism-driven biomarkers to guide immune checkpoint blockade in cancer therapy. Nat Rev Cancer 2016; 16: 275-287.
- [83] Simeone E, Grimaldi AM and Ascierto PA. Anti-PD1 and anti-PD-L1 in the treatment of meta-static melanoma. Melanoma Manag 2015; 2: 41-50.
- [84] Ciccarese C, Iacovelli R, Bria E, Modena A, Massari F, Brunelli M, Fantinel E, Bimbatti D, Zamboni GA, Artibani W and Tortora G. The incidence and relative risk of pulmonary toxicity in patients treated with anti-PD1/PD-L1 therapy for solid tumors: a meta-analysis of current studies. Immunotherapy 2017; 9: 579-587.

- [85] Alsaab HO, Sau S, Alzhrani R, Tatiparti K, Bhise K, Kashaw SK and Iyer AK. PD-1 and PD-L1 checkpoint signaling inhibition for cancer immunotherapy: mechanism, combinations, and clinical outcome. Front Pharmacol 2017; 8: 561.
- [86] Scott IJ. Nivolumab: a review in advanced melanoma. Drugs 2015; 75: 1413-1424.
- [87] Peters S, Kerr KM and Stahel R. PD-1 blockade in advanced NSCLC: a focus on pembrolizumab. Cancer Treat Rev 2018; 62: 39-49.
- [88] Zwaenepoel K, Jacobs J, De Meulenaere A, Silence K, Smits E, Siozopoulou V, Hauben E, Rolfo C, Rottey S and Pauwels P. CD70 and PD-L1 in anaplastic thyroid cancer-promising targets for immunotherapy. Histopathology 2017; 71: 357-365.
- [89] Cantara S, Bertelli E, Occhini R, Regoli M, Brilli L, Pacini F, Castagna MG and Toti P. Blockade of the programmed death ligand 1 (PD-L1) as potential therapy for anaplastic thyroid cancer. Endocrine 2019; 64: 122-129.
- [90] Goodman AM, Piccioni D, Kato S, Boichard A, Wang HY, Frampton G, Lippman SM, Connelly C, Fabrizio D, Miller V, Sicklick JK and Kurzrock R. Prevalence of PDL1 amplification and preliminary response to immune checkpoint blockade in solid tumors. JAMA Oncol 2018; 4: 1237-1244.
- [91] Brauner E, Gunda V, Vanden Borre P, Zurakowski D, Kim YS, Dennett KV, Amin S, Freeman GJ and Parangi S. Combining BRAF inhibitor and anti PD-L1 antibody dramatically improves tumor regression and anti tumor immunity in an immunocompetent murine model of anaplastic thyroid cancer. Oncotarget 2016; 7: 17194-17211.
- [92] Kollipara R, Schneider B, Radovich M, Babu S and Kiel PJ. Exceptional response with immunotherapy in a patient with anaplastic thyroid cancer. Oncologist 2017; 22: 1149-1151.
- [93] Aghajani MJ, Cooper A, McGuire H, Jeffries T, Saab J, Ismail K, de Souza P, Bray V, Fazekas de St Groth B, Niles N and Roberts TL. Pembrolizumab for anaplastic thyroid cancer: a case study. Cancer Immunol Immunother 2019; 68: 1921-1934.
- [94] Yin M, Di G and Bian M. Dysfunction of natural killer cells mediated by PD-1 and Tim-3 pathway in anaplastic thyroid cancer. Int Immunopharmacol 2018; 64: 333-339.
- [95] Carosella ED, Ploussard G, LeMaoult J and Desgrandchamps F. A systematic review of immunotherapy in urologic cancer: evolving roles for targeting of CTLA-4, PD-1/PD-L1, and HLA-G. Eur Urol 2015; 68: 267-279.
- [96] Robert C, Thomas L, Bondarenko I, O'Day S, Weber J, Garbe C, Lebbe C, Baurain JF, Testori A, Grob JJ, Davidson N, Richards J, Maio M, Hauschild A, Miller WH Jr, Gascon P, Lotem M,

- Harmankaya K, Ibrahim R, Francis S, Chen TT, Humphrey R, Hoos A and Wolchok JD. Ipilimumab plus dacarbazine for previously untreated metastatic melanoma. N Engl J Med 2011; 364: 2517-2526.
- [97] Eggermont AM, Chiarion-Sileni V, Grob JJ, Dummer R, Wolchok JD, Schmidt H, Hamid O, Robert C, Ascierto PA, Richards JM, Lebbe C, Ferraresi V, Smylie M, Weber JS, Maio M, Konto C, Hoos A, de Pril V, Gurunath RK, de Schaetzen G, Suciu S and Testori A. Adjuvant ipilimumab versus placebo after complete resection of high-risk stage III melanoma (EORTC 18071): a randomised, double-blind, phase 3 trial. Lancet Oncol 2015; 16: 522-530.
- [98] Tuccilli C, Baldini E, Sorrenti S, Catania A, Antonelli A, Fallahi P, Tartaglia F, Barollo S, Mian C, Palmieri A, Carbotta G, Arcieri S, Pironi D, Vergine M, Monti M and Ulisse S. CTLA-4 and PD-1 ligand gene expression in epithelial thyroid cancers. Int J Endocrinol 2018; 2018: 1742951.
- [99] Snyder A, Makarov V, Merghoub T, Yuan J, Zaretsky JM, Desrichard A, Walsh LA, Postow MA, Wong P, Ho TS, Hollmann TJ, Bruggeman C, Kannan K, Li Y, Elipenahli C, Liu C, Harbison CT, Wang L, Ribas A, Wolchok JD and Chan TA. Genetic basis for clinical response to CTLA-4 blockade in melanoma. N Engl J Med 2014; 371: 2189-2199.
- [100] Rizvi NA, Hellmann MD, Snyder A, Kvistborg P, Makarov V, Havel JJ, Lee W, Yuan J, Wong P, Ho TS, Miller ML, Rekhtman N, Moreira AL, Ibrahim F, Bruggeman C, Gasmi B, Zappasodi R, Maeda Y, Sander C, Garon EB, Merghoub T, Wolchok JD, Schumacher TN and Chan TA. Cancer immunology. Mutational landscape determines sensitivity to PD-1 blockade in nonsmall cell lung cancer. Science 2015; 348: 124-128.
- [101] Pathria P, Louis TL and Varner JA. Targeting tumor-associated macrophages in cancer. Trends Immunol 2019; 40: 310-327.
- [102] Kumar V, Donthireddy L, Marvel D, Condamine T, Wang F, Lavilla-Alonso S, Hashimoto A, Vonteddu P, Behera R, Goins MA, Mulligan C, Nam B, Hockstein N, Denstman F, Shakamuri S, Speicher DW, Weeraratna AT, Chao T, Vonderheide RH, Languino LR, Ordentlich P, Liu Q, Xu X, Lo A, Pure E, Zhang C, Loboda A, Sepulveda MA, Snyder LA and Gabrilovich DI. Cancer-associated fibroblasts neutralize the antitumor effect of CSF1 receptor blockade by inducing PMN-MDSC infiltration of tumors. Cancer Cell 2017; 32: 654-668, e655.
- [103] French JD. Revisiting immune-based therapies for aggressive follicular cell-derived thyroid cancers. Thyroid 2013; 23: 529-542.

- [104] Weber F, Junger H, Werner JM, Velez Char N, Rejas C, Schlitt HJ and Hornung M. Increased cytoplasmatic expression of cancer immune surveillance receptor CD1d in anaplastic thyroid carcinomas. Cancer Med 2019; 8: 7065-7073.
- [105] Schurch CM, Roelli MA, Forster S, Wasmer MH, Bruhl F, Maire RS, Di Pancrazio S, Ruepp MD, Giger R, Perren A, Schmitt AM, Krebs P, Charles RP and Dettmer MS. Targeting CD47 in anaplastic thyroid carcinoma enhances tumor phagocytosis by macrophages and is a promising therapeutic strategy. Thyroid 2019; 29: 979-992.
- [106] Zaretsky JM, Garcia-Diaz A, Shin DS, Escuin-Ordinas H, Hugo W, Hu-Lieskovan S, Torrejon DY, Abril-Rodriguez G, Sandoval S, Barthly L, Saco J, Homet Moreno B, Mezzadra R, Chmielowski B, Ruchalski K, Shintaku IP, Sanchez PJ, Puig-Saus C, Cherry G, Seja E, Kong X, Pang J, Berent-Maoz B, Comin-Anduix B, Graeber TG, Tumeh PC, Schumacher TN, Lo RS and Ribas A. Mutations associated with acquired resistance to PD-1 blockade in melanoma. N Engl J Med 2016; 375: 819-829.
- [107] Gao J, Shi LZ, Zhao H, Chen J, Xiong L, He Q, Chen T, Roszik J, Bernatchez C, Woodman SE, Chen PL, Hwu P, Allison JP, Futreal A, Wargo JA and Sharma P. Loss of IFN-gamma pathway genes in tumor cells as a mechanism of resistance to anti-CTLA-4 therapy. Cell 2016; 167: 397-404, e399.
- [108] Chintakuntlawar AV, Yin J, Foote RL, Kasperbauer JL, Rivera M, Asmus E, Garces NI, Janus JR, Liu M, Ma DJ, Moore EJ, Morris JC 3rd, Neben-Wittich M, Price DL, Price KA, Ryder M, Van Abel KM, Hilger C, Samb E and Bible KC. A phase 2 study of pembrolizumab combined with chemoradiotherapy as initial treatment for anaplastic thyroid cancer. Thyroid 2019; 29: 1615-1622.
- [109] Rotte A, Jin JY and Lemaire V. Mechanistic overview of immune checkpoints to support the rational design of their combinations in cancer immunotherapy. Ann Oncol 2018; 29: 71-83.
- [110] Ishizuka JJ, Manguso RT, Cheruiyot CK, Bi K, Panda A, Iracheta-Vellve A, Miller BC, Du PP, Yates KB, Dubrot J, Buchumenski I, Comstock DE, Brown FD, Ayer A, Kohnle IC, Pope HW, Zimmer MD, Sen DR, Lane-Reticker SK, Robitschek EJ, Griffin GK, Collins NB, Long AH, Doench JG, Kozono D, Levanon EY and Haining WN. Loss of ADAR1 in tumours overcomes resistance to immune checkpoint blockade. Nature 2019; 565: 43-48.